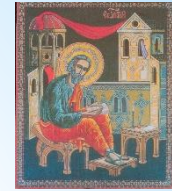




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Почетен член на "Съвета на Европейската научна и културна общност" БСТ



Interactive System for Education in Modelling and Control of Bioprocesses

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Anastasiya Zlatkova, Ph.D.

Denitsa Kristeva

May 2022
Sibiu, Roumania

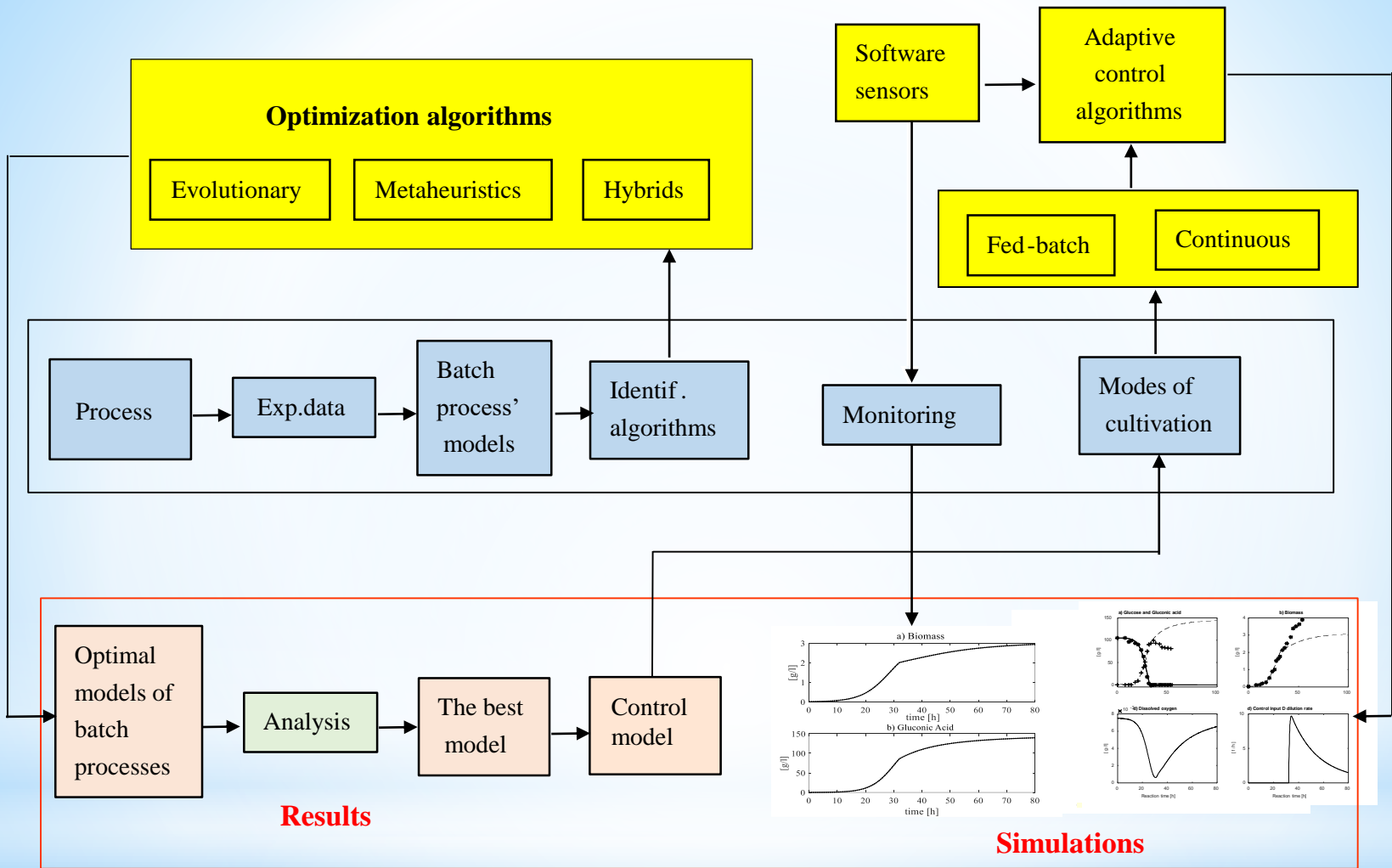


Fig. 1. Open source system InSEMCoBio

Setting up a fermentation process model and metaheuristic algorithm parameters in InSEMCoBio

Identification Panel

Current Step

- Select Fermentation Process
- Select Model and Kinetics
- Load Experimental Data
- Model Parameter Identification

Choose Fermentation Process

E. coli MC4110 Fed-batch

Choose Model and Kinetics

Mass Balance Equations

- $dX/dt = \mu \cdot X - F/V \cdot X$
- $dS/dt = -1/Y_{xs} \cdot \mu \cdot X + (S_0 - S) \cdot F/V$
- $dO_2/dt = 1/Y_{ox} \cdot \mu \cdot X + K_{la} \cdot (O_2^* - O_2) - F/V \cdot O_2$
- $dV/dt = F$

Kinetic Models

- Monod
- Contoa
- Andrew

Set Model Load Data

Choose Algorithm

Genetic Algorithm

Set Algorithm Parameters

MUTR (0.001, 0.1)

XOVR (0.1, 1.0)

NIND (1, 200)

MAXGEN (1, 400)

GGAP (0.1, 1)

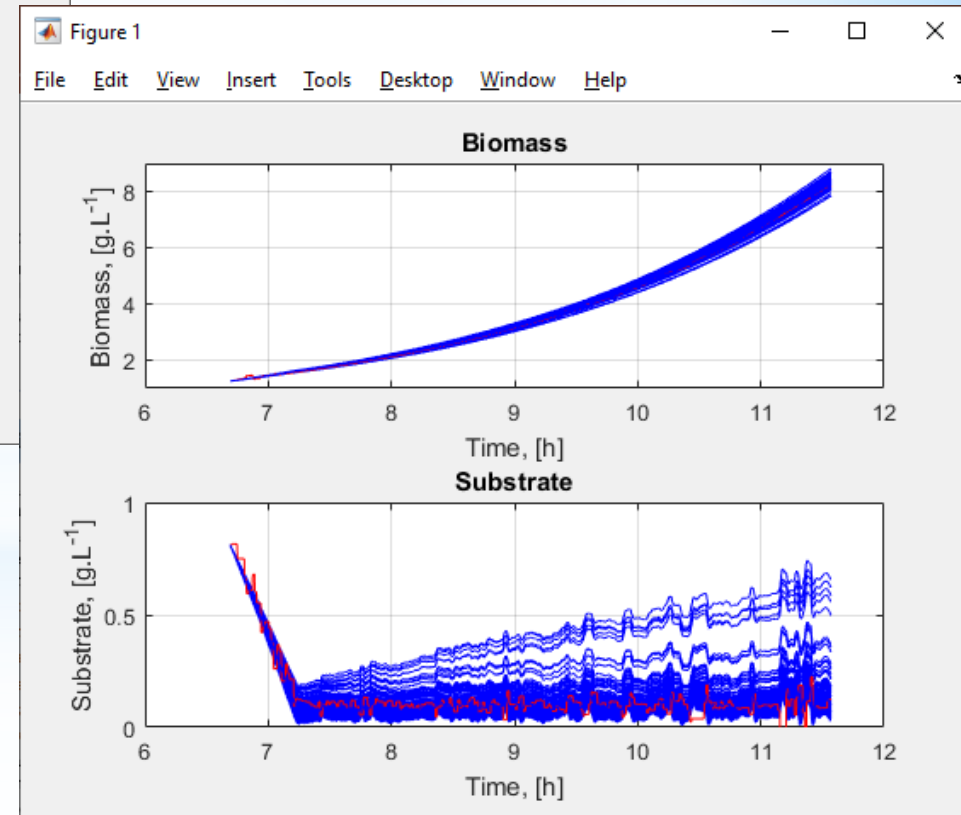
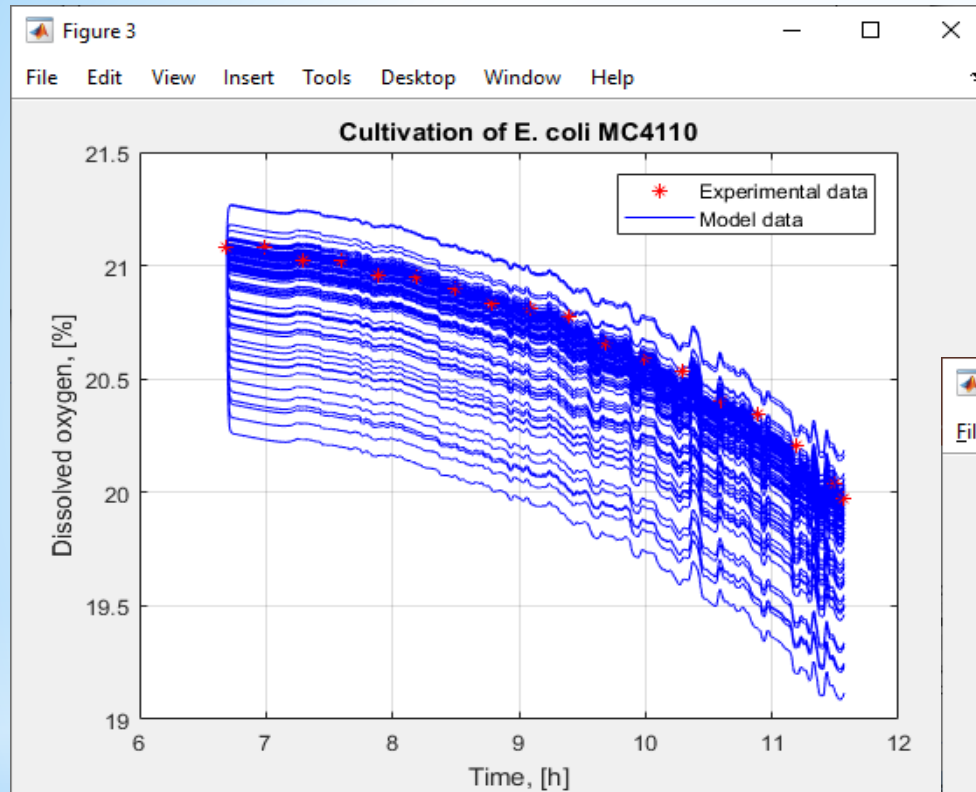
Run Plot Results

Logs

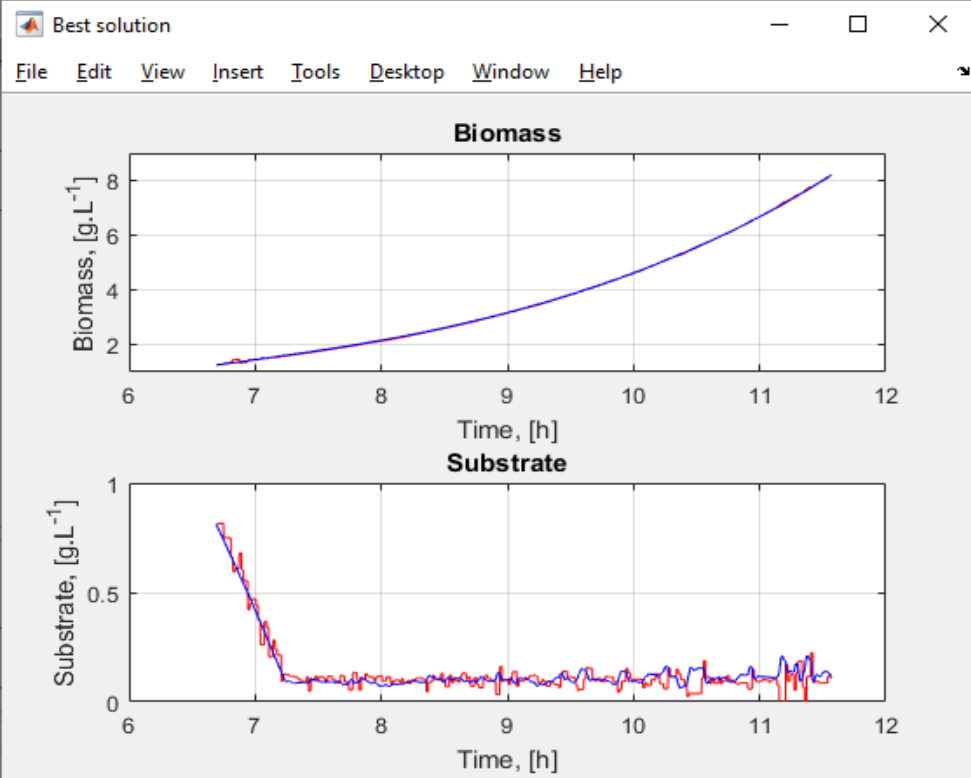
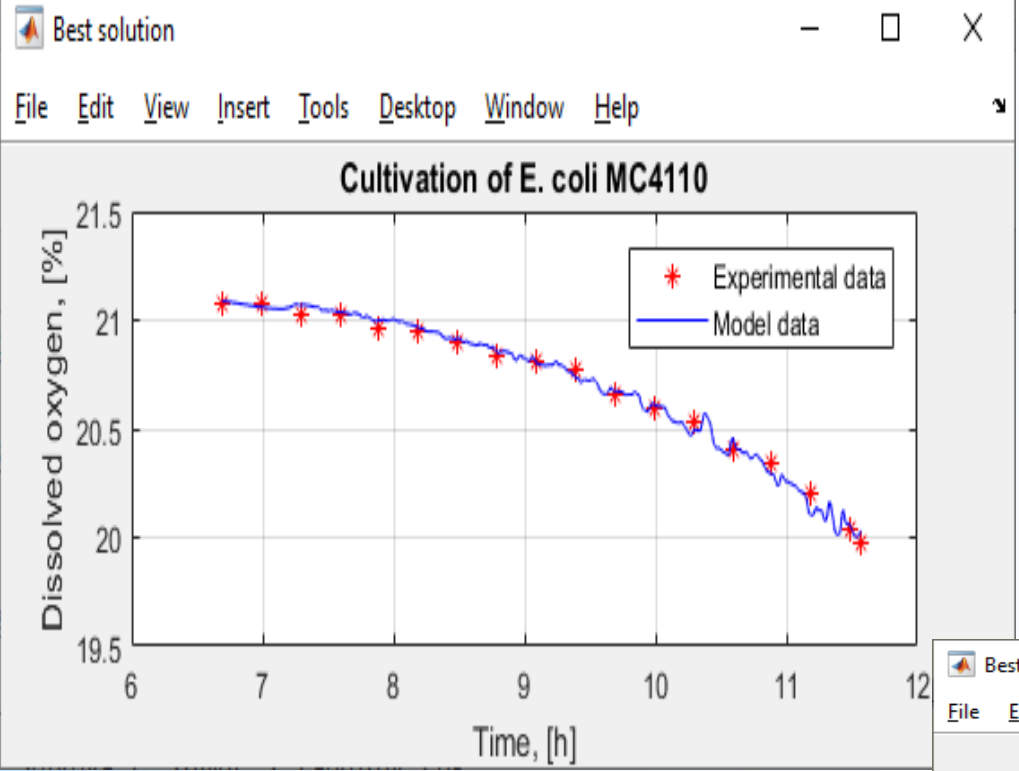
Step	Record
FP	E. coli MC4110 Fed-batch
Data	EcoliDataSet.xls

MKA Results

Visualizing the results of the identification procedure in InSEMCoBio

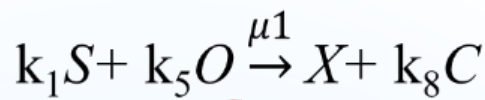


Visualizing the results of the identification procedure in InSEMCoBio



Model Identification and Monitoring of E. Coli fed-batch cultivation for extracellular production of bacterial phytase

Oxidative-fermentative growth model on glucose



Oxidative growth on glucose

Model

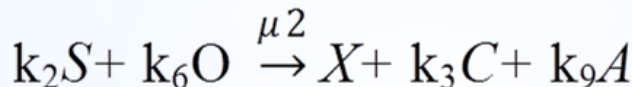
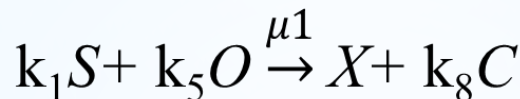
$$\frac{d}{dt} \begin{bmatrix} X \\ S \\ A \\ O \\ C_r \end{bmatrix} = \begin{bmatrix} 1 \\ -k_1 \\ 0 \\ -k_5 \\ k_8 \end{bmatrix} \mu_1 X - D \begin{bmatrix} X \\ S \\ A \\ O \\ C_r \end{bmatrix} + \frac{F_{in,S}}{W} \begin{bmatrix} 0 \\ S_{in} \\ 0 \\ 0 \\ 0 \end{bmatrix} + \begin{bmatrix} 0 \\ 0 \\ 0 \\ OTR \\ -CTR \end{bmatrix}$$

$$q_s = q_{s,max} S / (K_s + S)$$

$$\mu_1 = q_s / k_1$$

Marker

$$R_{ac} = \frac{dA}{dt} + \frac{F_{in,s}}{W} A$$



Oxidative-fermentative growth on glucose

Model

$$R_{ac} > 0$$

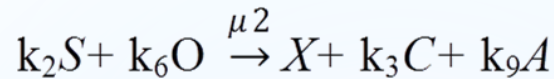
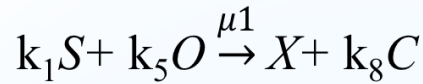
$$\frac{d}{dt} \begin{bmatrix} X \\ S \\ A \\ O \\ C_r \end{bmatrix} = \begin{bmatrix} 1 & 1 \\ -k_1 & -k_2 \\ 0 & k_3 \\ -k_5 & -k_6 \\ k_8 & k_9 \end{bmatrix} \begin{bmatrix} \mu_1 \\ \mu_2 \end{bmatrix} X - D \begin{bmatrix} X \\ S \\ A \\ O \\ C_r \end{bmatrix} + \frac{F_{in,S}}{W} \begin{bmatrix} 0 \\ S_{in} \\ 0 \\ 0 \\ 0 \end{bmatrix} + \begin{bmatrix} 0 \\ 0 \\ 0 \\ OTR \\ -CTR \end{bmatrix}$$

$$\mu_1 = q_{s,crit} / k_1$$

$$\mu_2 = (q_s - q_{s,crit}) / k_2$$

$$q_{s,crit} = \frac{q_{o,max}}{k_{os}} \frac{K_{i,o}}{K_{i,o} + A}$$

Oxidative-fermentative growth model on glucose and oxidative on acetate



Oxidative-fermentative growth on glucose

Model

$$R_{ac} > 0$$



$$\frac{d}{dt} \begin{bmatrix} X \\ S \\ A \\ O \\ C_r \end{bmatrix} = \begin{bmatrix} 1 & 1 \\ -k_1 & -k_2 \\ 0 & k_3 \\ -k_5 & -k_6 \\ k_8 & k_9 \end{bmatrix} \begin{bmatrix} \mu_1 \\ \mu_2 \end{bmatrix} X - D \begin{bmatrix} X \\ S \\ A \\ O \\ C_r \end{bmatrix} + \frac{F_{in,S}}{W} \begin{bmatrix} 0 \\ S_{in} \\ 0 \\ 0 \\ 0 \end{bmatrix} + \begin{bmatrix} 0 \\ 0 \\ 0 \\ OTR \\ -CTR \end{bmatrix}$$

$$\mu_1 = q_{s,crit} / k_1$$

$$\mu_2 = (q_s - q_{s,crit}) / k_2$$

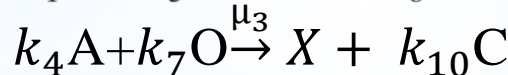
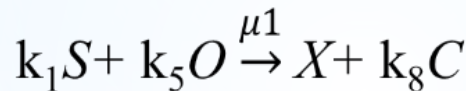
$$q_{s,crit} = \frac{q_{o,max}}{k_{os}} \frac{K_{i,o}}{K_{i,o} + A}$$

Marker

$$R_{ac} = \frac{dA}{dt} + \frac{F_{in,s}}{W} A$$



$$R_{ac} < 0$$



Oxidative growth on glucose and acetate

Model

$$\frac{d}{dt} \begin{bmatrix} X \\ S \\ A \\ O \\ C_r \end{bmatrix} = \begin{bmatrix} 1 & 1 \\ -k_1 & 0 \\ 0 & -k_4 \\ -k_5 & -k_7 \\ k_8 & k_{10} \end{bmatrix} \begin{bmatrix} \mu_1 \\ \mu_3 \end{bmatrix} X - D \begin{bmatrix} X \\ S \\ A \\ O \\ C_r \end{bmatrix} + \frac{F_{in,S}}{W} \begin{bmatrix} 0 \\ S_{in} \\ 0 \\ 0 \\ 0 \end{bmatrix} + \begin{bmatrix} 0 \\ 0 \\ 0 \\ OTR \\ -CTR \end{bmatrix}$$

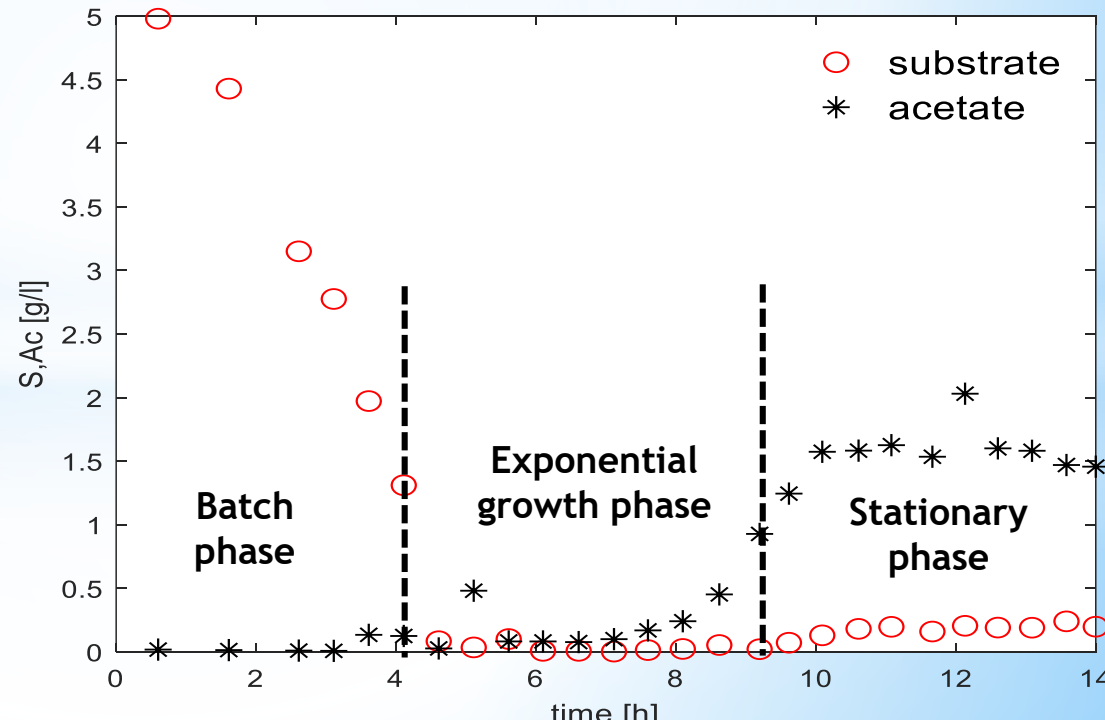
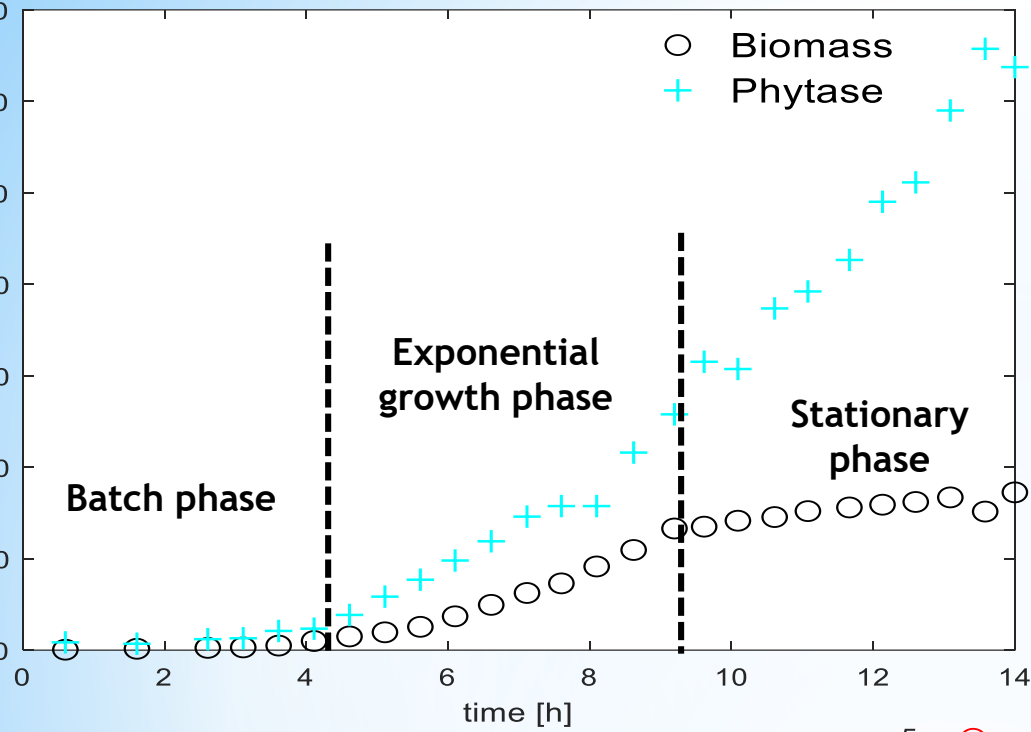
$$\mu_1 = q_{s,crit} / k_1$$

$$\mu_3 = q_{ac} / k_4$$

$$q_{s,crit} = \frac{q_{o,max}}{k_{os}} \frac{K_{i,o}}{K_{i,o} + A}$$

$$q_{ac} = q_{ac,max} \left(\frac{A}{K_A + A} \right) \left(\frac{K_{i,A}}{K_{i,A} + A} \right)$$

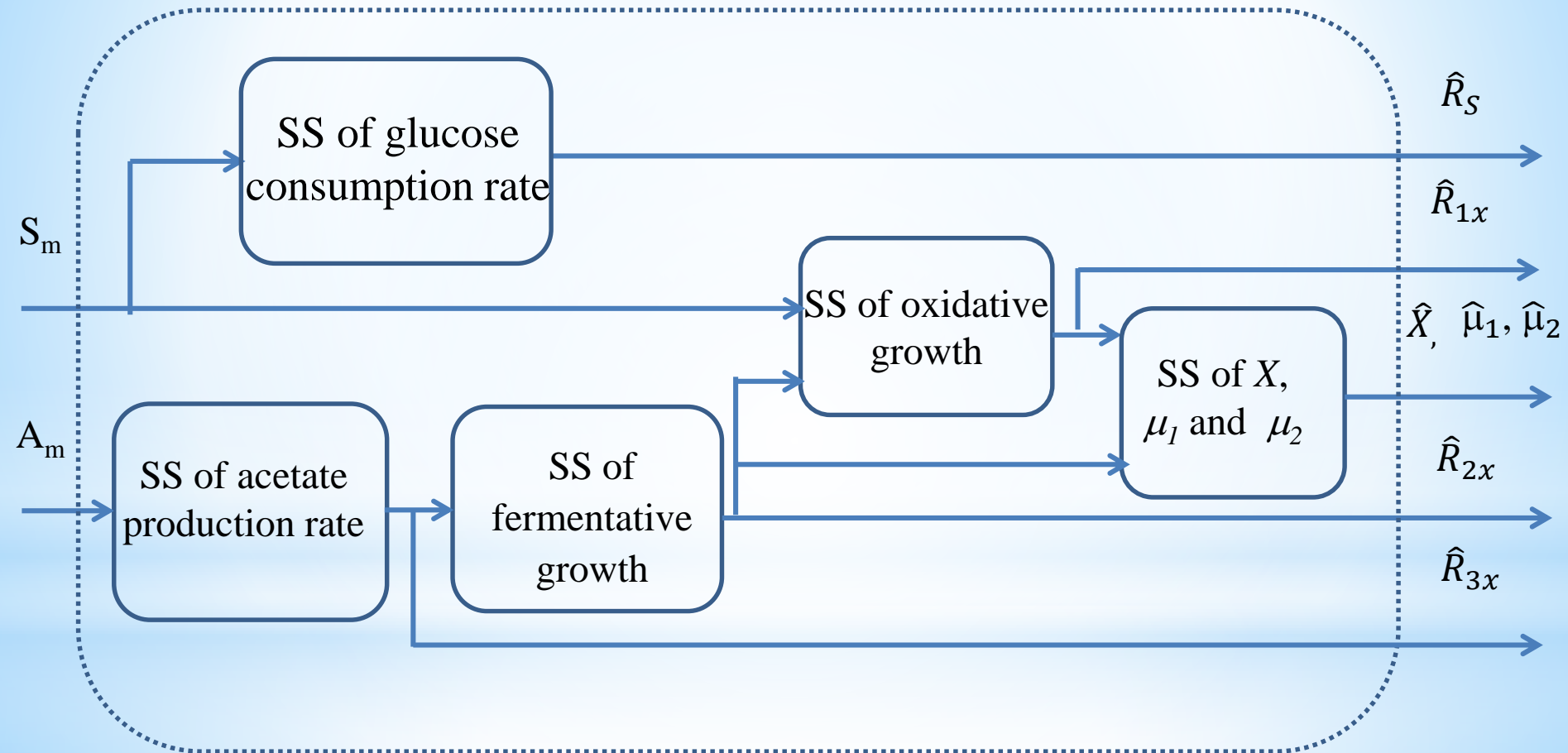
Experimental data



Comparison of the coefficients of the best models for the three phases

	q_{smax}	k_s	k_{is}	q_{lomax}	k_{os}	k_{io}	q_{acmax}	k_a	k_{ia}	k_1	k_2	k_3	k_4	k_5	k_6	k_7
1 phase	4.19	0.19	5.54	1.1	2.15	0.088	0.082	1.17	-	3.69	0.557	0.187	4.6	1.41	2.66	0.45
2 phase	34.24	0.79	1.83	0.469	2.53	0.197	0.143	0.97	0.246	2.08	2.167	0.049	4.1	2.88	1.52	0.5
3 phase	77.11	0.47	12.3	2.1	3.29	0.134	0.0021	0.295	0.228	16.6	11.66	0.42	9.9	39.45	9.53	0.56

Scheme of the cascade software sensor of the process kinetics



On-line estimation of acetate production and consumption rates

Acetate production

$$\frac{d\hat{A}}{dt} = \hat{R}_{ap} - DA_m + w_1(A_m - \hat{A})$$

$$\frac{d\hat{R}_{ap}}{dt} = w_2(A_m - \hat{A})$$

Acetate consumption

$$\frac{d\hat{A}}{dt} = -\hat{R}_{ac} - DA_m + \lambda_1(A_m - \hat{A})$$

$$\frac{d\hat{R}_{ac}}{dt} = -\lambda_2(A_m - \hat{A})$$

On-line estimation of R_{1X} , R_{2X} , R_{3X} and X

$$\frac{d\hat{S}}{dt} = -k_1\hat{R}_{1X} - k_2\hat{R}_{ap}/k_3 - \frac{F}{V}S_m + \frac{F_{in,S}}{V}S_{in} + \lambda_1(S_m - \hat{S})$$

$$\frac{d\hat{R}_{1X}}{dt} = \lambda_2(S_m - \hat{S})$$

$$\hat{R}_{2X} = \hat{R}_{ap}/k_3$$

$$\hat{\mu}_1 = \hat{R}_{1X}/\hat{X}$$

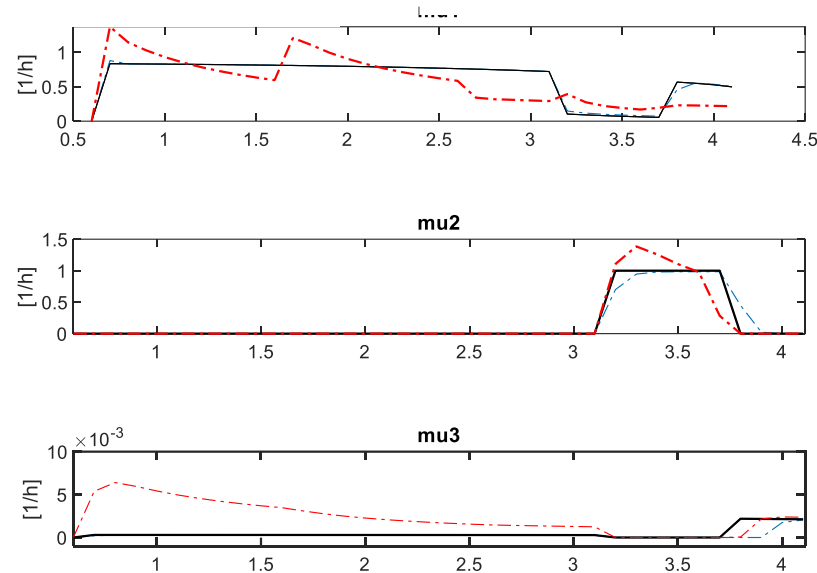
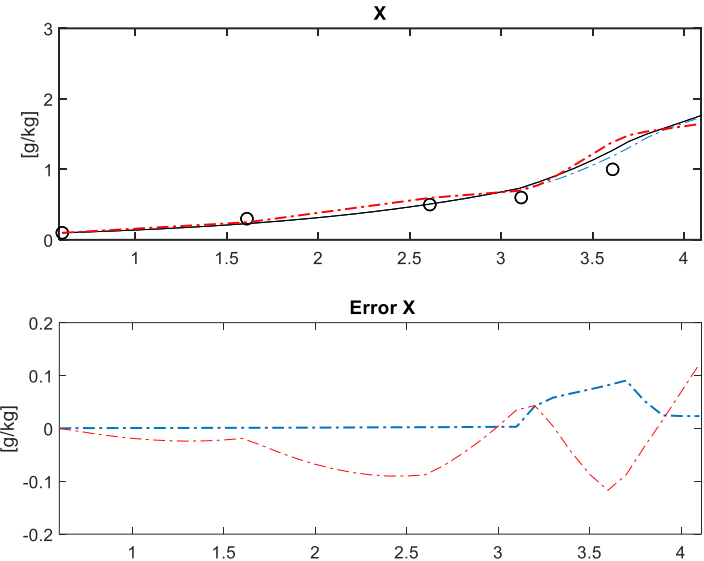
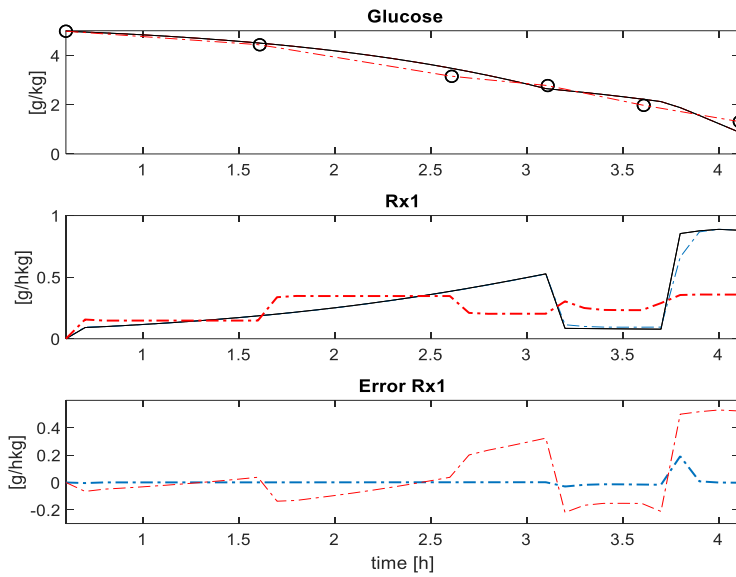
$$\hat{R}_{3X} = -\hat{R}_{ac}/k_4$$

$$\hat{\mu}_2 = \hat{R}_{2X}/\hat{X}$$

$$\frac{d\hat{X}}{dt} = \hat{R}_{1X} + \hat{R}_{2X} + \hat{R}_{3X} - D\hat{X}$$

$$\hat{\mu}_3 = \hat{R}_{3X}/\hat{X}$$

Monitoring results– I phase



Tuning parameters

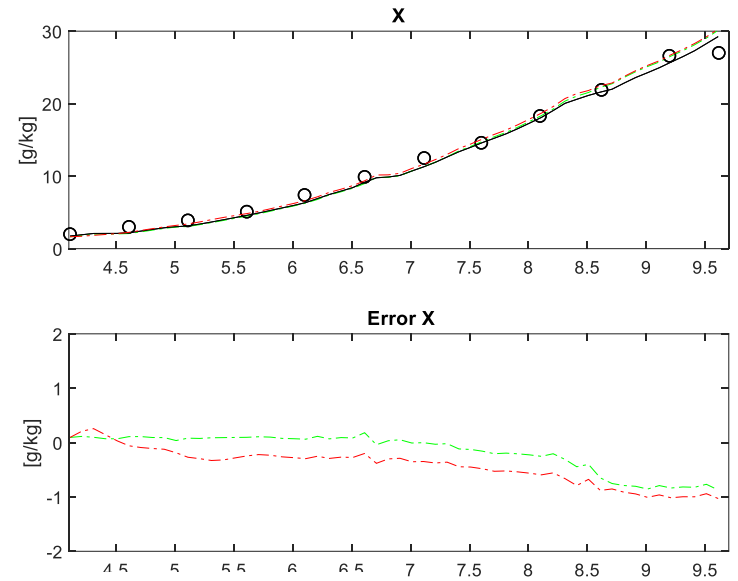
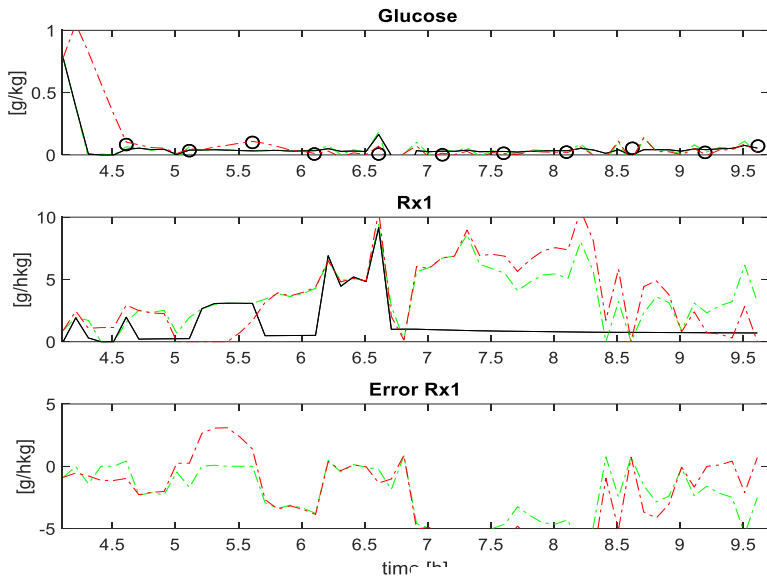
$C1=50;$

$C2=50;$

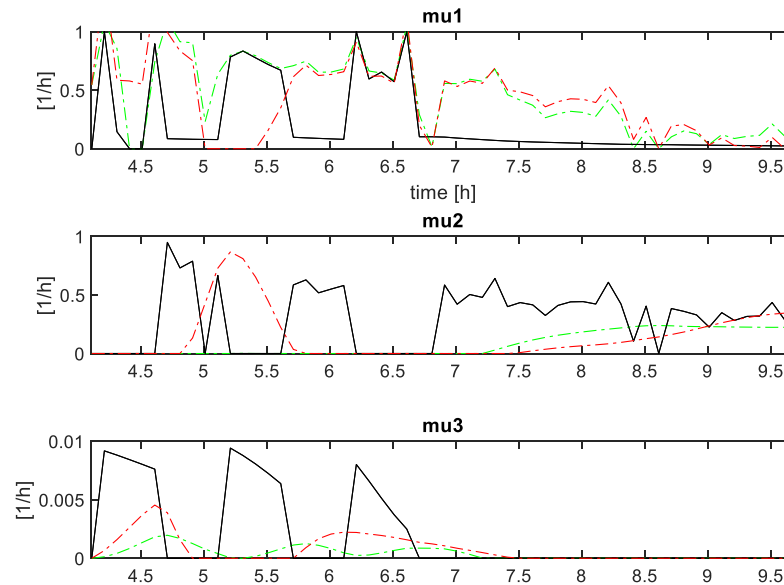
Black lines- model data

SS results: with red lines - inputs are experimental data, with blue lines – inputs are model data

Monitoring results– II phase

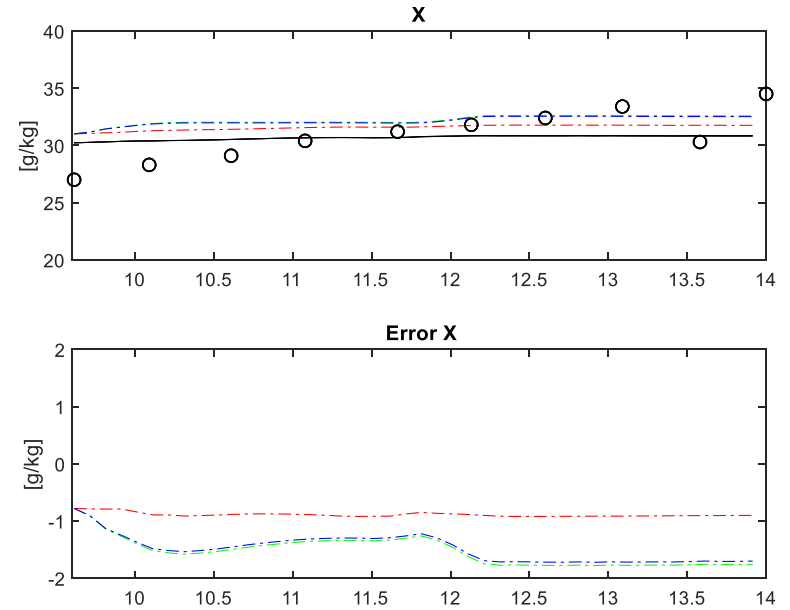
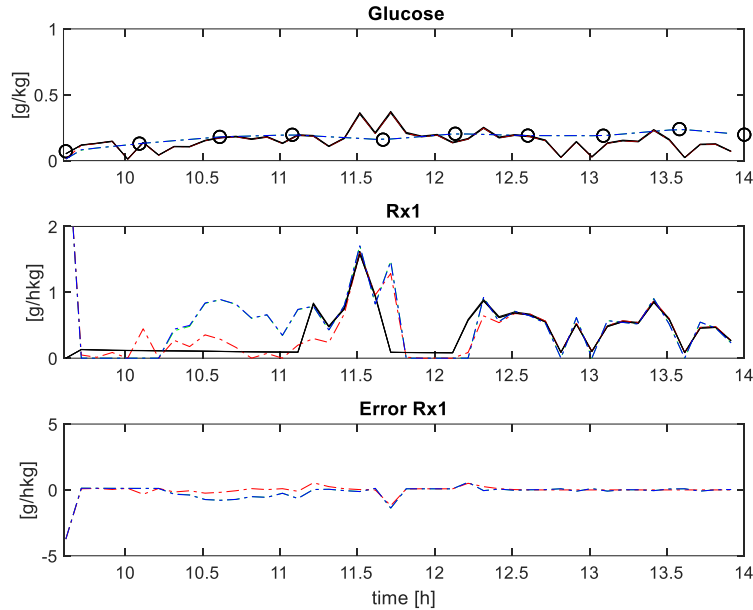


SS tuning
parameters
C1= 54.7764;
C2= 2.7926;
Black lines- model
data



SS results: with red lines - inputs are experimental data, with green lines – inputs are model data

Monitoring results– III phase



SS tuning parameters

$C1=77.0215$

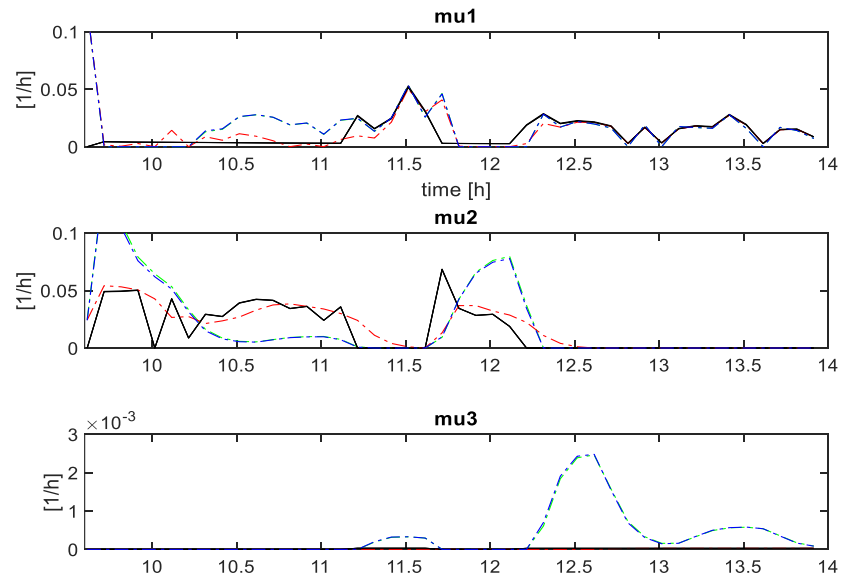
$C2=25$

red lines – model data

green lines ($C2=1.3$) и blue

lines ($C2=25$)

– experimental data

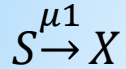


Comparison of the coefficients of the best models for the three phases

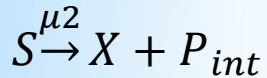
	q_{smax}	k_s	k_{is}	q_{lmax}	k_{os}	k_{io}	q_{acmax}	k_a	k_{ia}	k_1	k_2	k_3	k_4	k_5	k_6	k_7
1 phase	4.19	0.19	5.54	1.1	2.15	0.088	0.082	1.17	-	3.69	0.557	0.187	4.6	1.41	2.66	0.45
2 phase	34.24	0.79	1.83	0.469	2.53	0.197	0.143	0.97	0.246	2.08	2.167	0.049	4.1	2.88	1.52	0.5
3 phase	77.11	0.47	12.3	2.1	3.29	0.134	0.0021	0.295	0.228	16.6	11.66	0.42	9.9	39.45	9.53	0.56

A Methodology for Simultaneously State and Kinetics Observation of a Class Controllable Bioprocesses

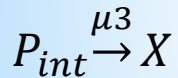
Oxidative growth on main carbon source, with specific growth rate μ_1 :



Fermentative growth on main carbon source, with specific growth rate μ_2 :



Oxidative growth on intermediate product, with specific growth rate μ_3 :



As can be seen from the last reaction, the main source of carbon is depleted and biomass increases at the expense of the intermediate. Its concentration is usually low, leading to insignificant growth of biomass and low productivity of the target product. To avoid this reaction, the processes are controlled by feeding on a basic carbon source.

This is done through two modes of control – fed-batch or continuous modes of cultivations. For the considered class processes is accepted that transport dynamics, main carbon source and intermediate product are on-line measured.

.

A Methodology for Simultaneously State and Kinetics Observation of a Class Bioprocesses

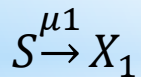
From what has been said so far, it is clear that the considered processes have two specific growth rates and total biomass.

The new idea is to present the biomass as consisting of two parts - the first one is connected with oxidative growth on main carbon source, X_1 , the second – with fermentative growth on main carbon source, X_2 . The full biomass concentration will be the sum of X_1 and X_2 :

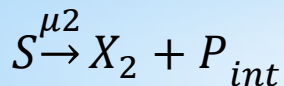
$$X = X_1 + X_2$$

The reaction scheme for the considered case is rewritten:

Oxidative growth on main carbon source, with specific growth rate μ_1 :



Fermentative growth on main carbon source, with specific growth rate μ_2 :



Estimator of oxidative growth of biomass X_1 with specific growth μ_1

$$\frac{dZ_1}{dt} = -\frac{F}{V}Z_1 + \frac{F}{V}S_{in}$$

$$\frac{d\hat{S}}{dt} = (S - Z_1)\hat{\mu}_1 - \frac{F}{V}(S - S_{in}) - k_2\hat{\mu}_2\hat{X}_2 + w_1(S - \hat{S})$$

$$\frac{d\hat{\mu}_1}{dt} = w_2(S - Z_1)(S - \hat{S})$$

$$\frac{dV}{dt} = F$$

Estimator of fermentative growth of biomass X_2 with specific growth μ_2

$$\frac{dZ_2}{dt} = -\frac{F}{V}Z_2$$

$$\frac{d\hat{P}_{int}}{dt} = (P_{int} - Z_2)\hat{\mu}_2 - \frac{F}{V}P_{int} + \gamma_1(P_{int} - \hat{P}_{int})$$

$$\frac{d\hat{\mu}_2}{dt} = \gamma_2(P_{int} - \hat{P}_{int})(P_{int} - Z_2)$$

$$\frac{dV}{dt} = F$$

Observers of biomasses X_1 and X_2

$$\frac{d}{dt} \begin{bmatrix} \hat{X}_1 \\ \hat{X}_2 \end{bmatrix} = \begin{bmatrix} \hat{\mu}_1 & 0 \\ 0 & \hat{\mu}_2 \end{bmatrix} \begin{bmatrix} \hat{X}_1 \\ \hat{X}_2 \end{bmatrix} - \frac{F}{V} \begin{bmatrix} \hat{X}_1 \\ \hat{X}_2 \end{bmatrix}$$

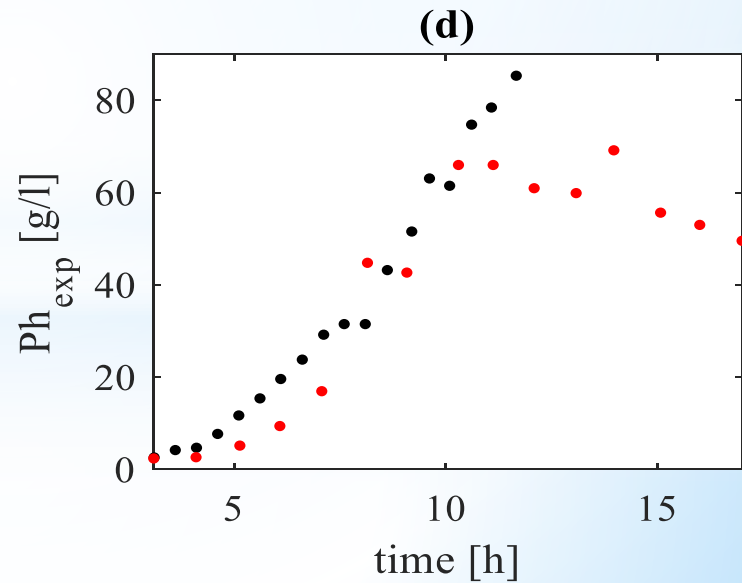
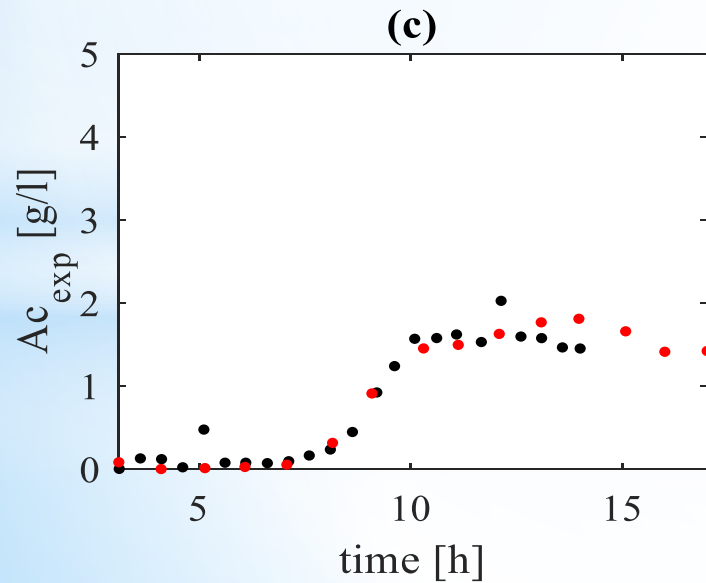
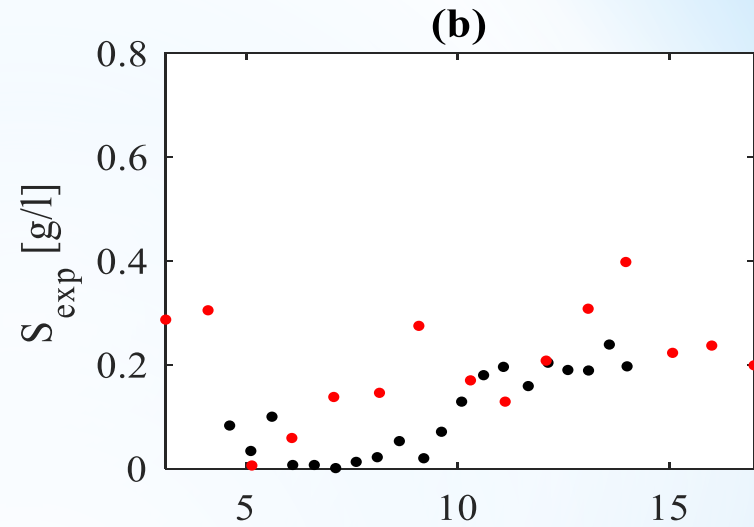
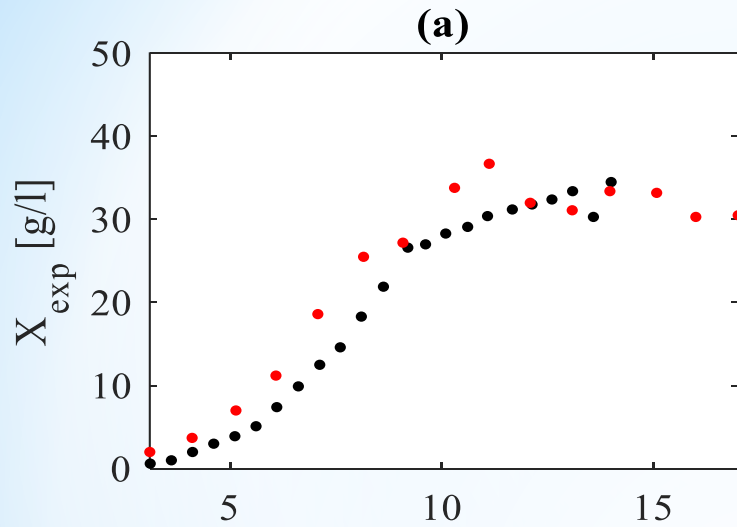
$$\frac{dV}{dt} = F$$

$$\hat{X} = \hat{X}_1 + \hat{X}_2$$

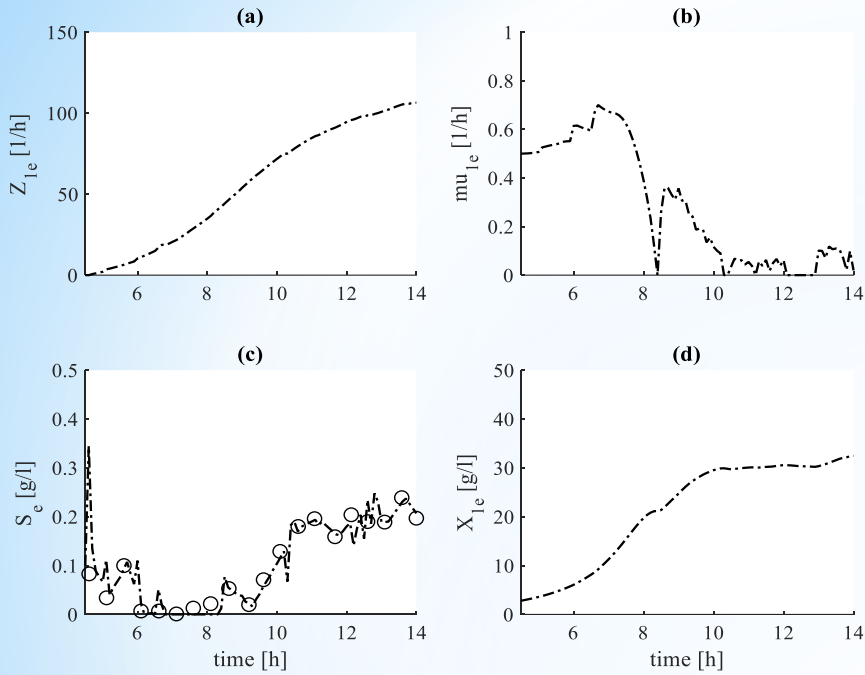
Observer of target product

$$\frac{d\hat{P}h}{dt} = k_{ph1}\hat{\mu}_1\hat{X}_1 + k_{ph2}\hat{\mu}_2\hat{X}_2 - \frac{F}{V}Ph$$

Two sets of experimental data

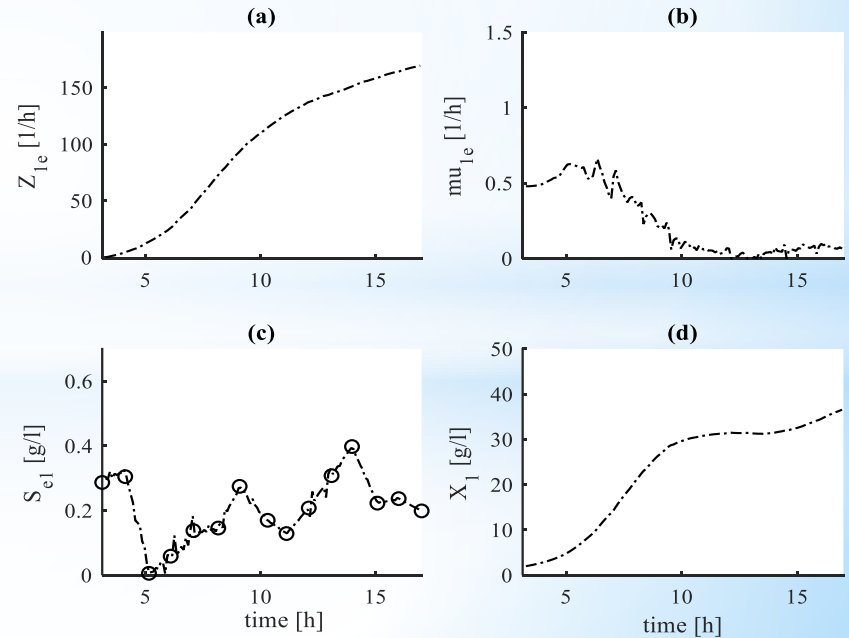


Estimates of μ_1 and X_1

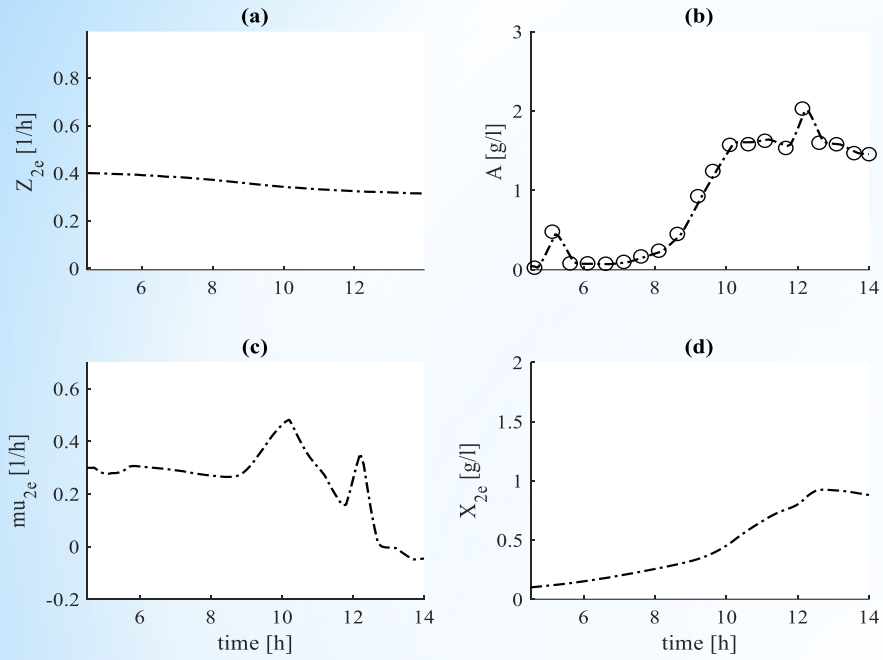


← Tuning

Verification →

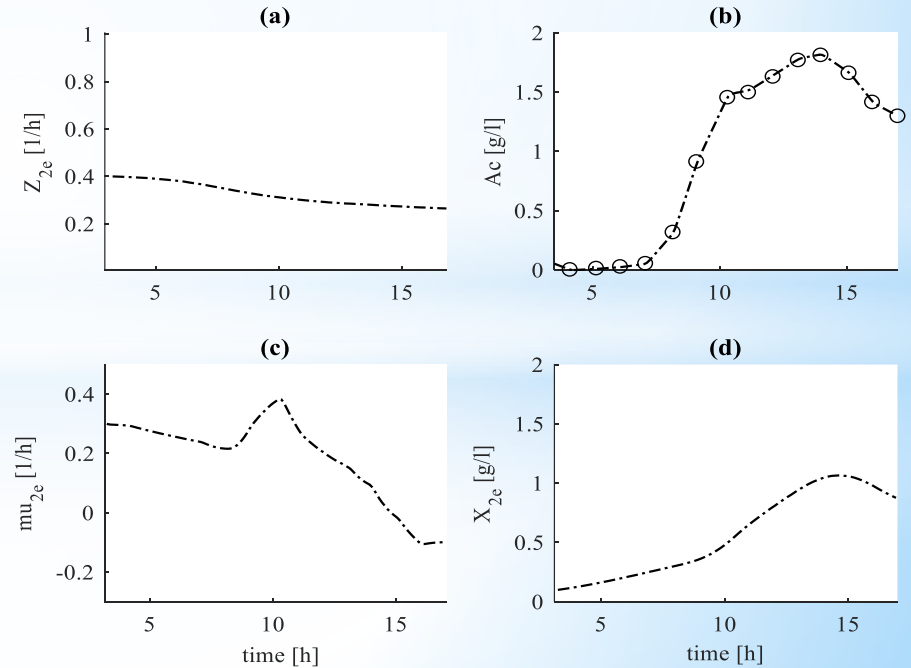


Estimates of μ_2 and X_2

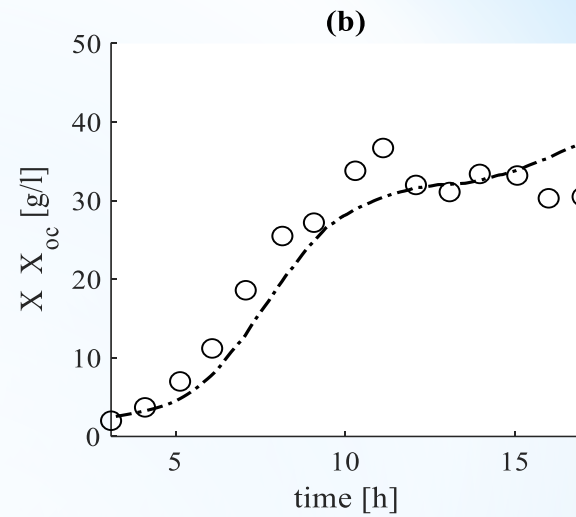
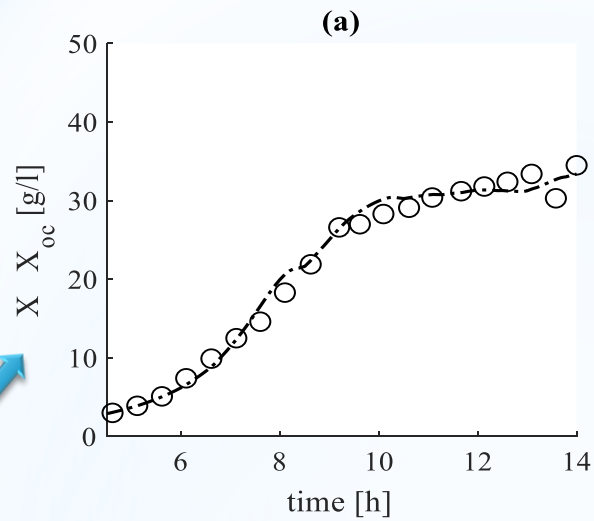


← Tuning

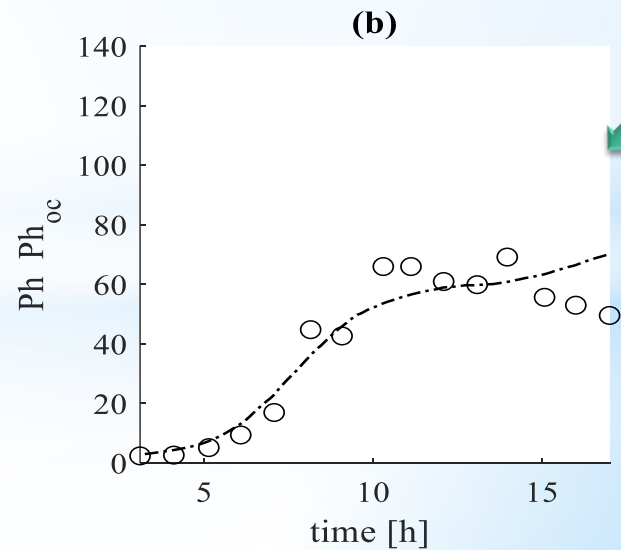
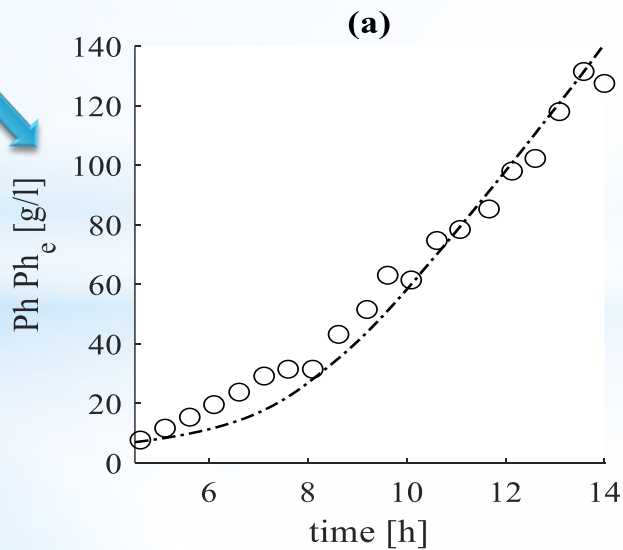
Verification →



Observers of X and Ph



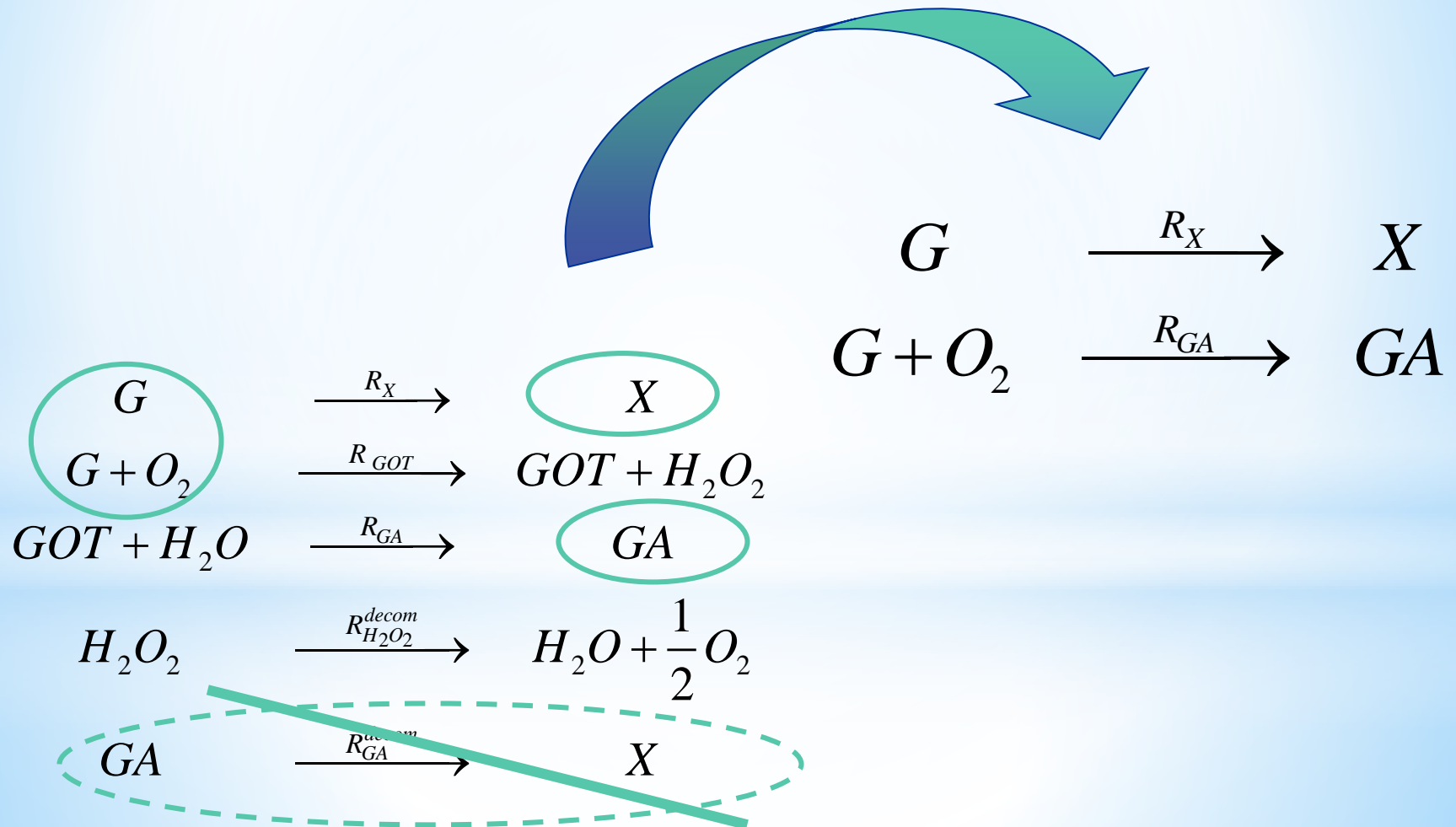
Tuning



Verification

*Model Identification, Monitoring and
Control of Aspergillus Niger
Fermentation for Gluconic Acid
Production*

Biochemical model and reaction scheme reduction



Reduced biochemical model

$$\frac{dX}{dt} = R_x;$$

$$\frac{dG}{dt} = -R_x - R_{GOT};$$

$$\frac{dGOT}{dt} = R_{GOT} - R_{GA};$$

$$\frac{dGA}{dt} = R_{GA};$$

$$\frac{dO}{dt} = -R_{GOT} + 0.5R_{H_2O_2} + K_L a(O_2^* - O_2);$$

$$\frac{dH_2O_2}{dt} = -R_{H_2O_2};$$

where

$$R_{H_2O_2} = R_{GOT} - R_{H_2O_2}^{decom};$$

$$R_x = \mu_{\max} X \frac{k - X}{k};$$

$$R_{GA} = \mu_{GA} GA \frac{(k_{GA} - GA)}{k_{GA}};$$

$$\frac{dX}{dt} = R_x;$$

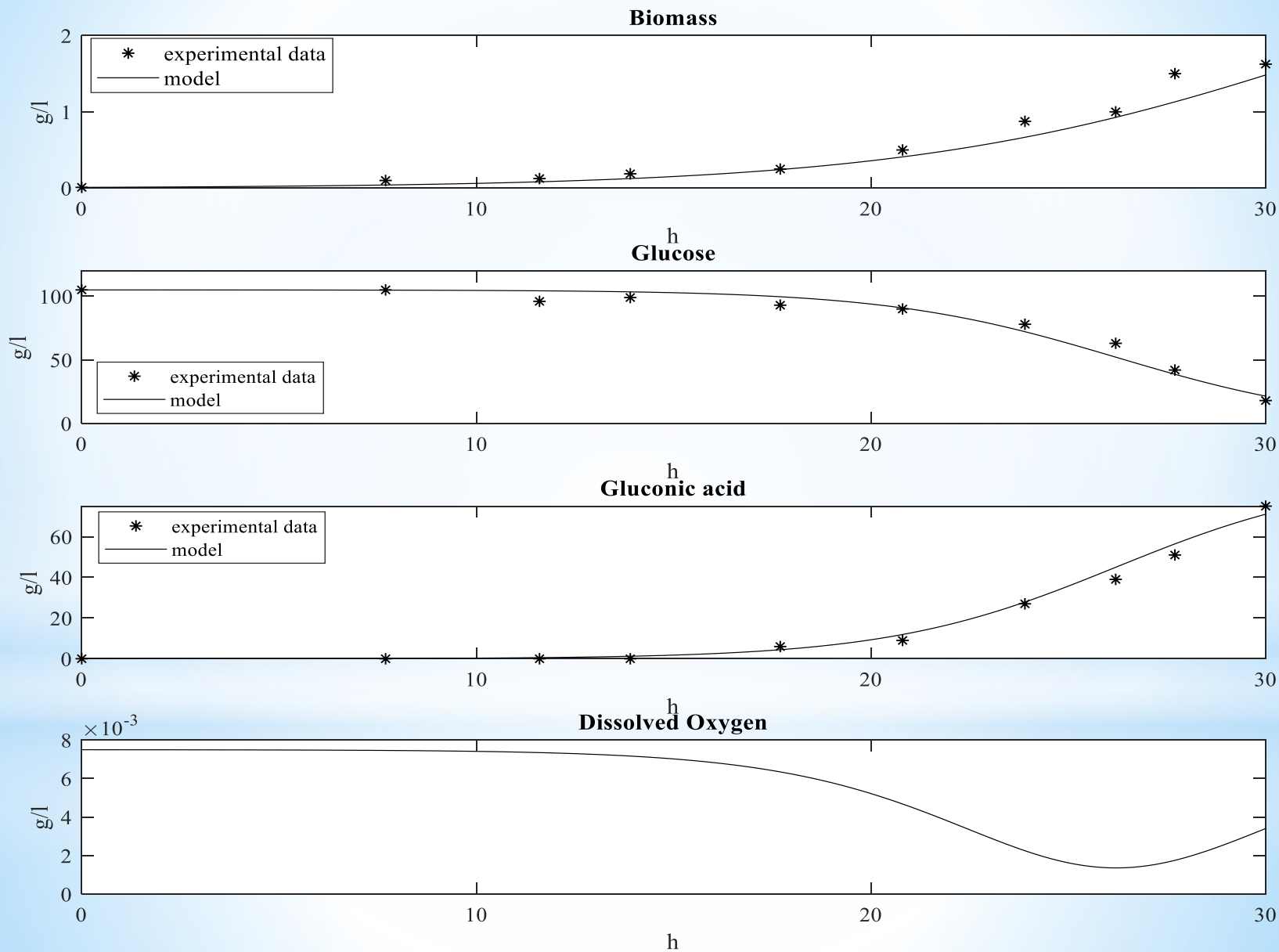
$$\frac{dG}{dt} = -R_x - R_{GA};$$

$$\frac{dGA}{dt} = R_{GA};$$

$$\frac{dO}{dt} = -0.5R_{GA} + K_L a(O_2^* - O_2),$$



Reduced model simulation

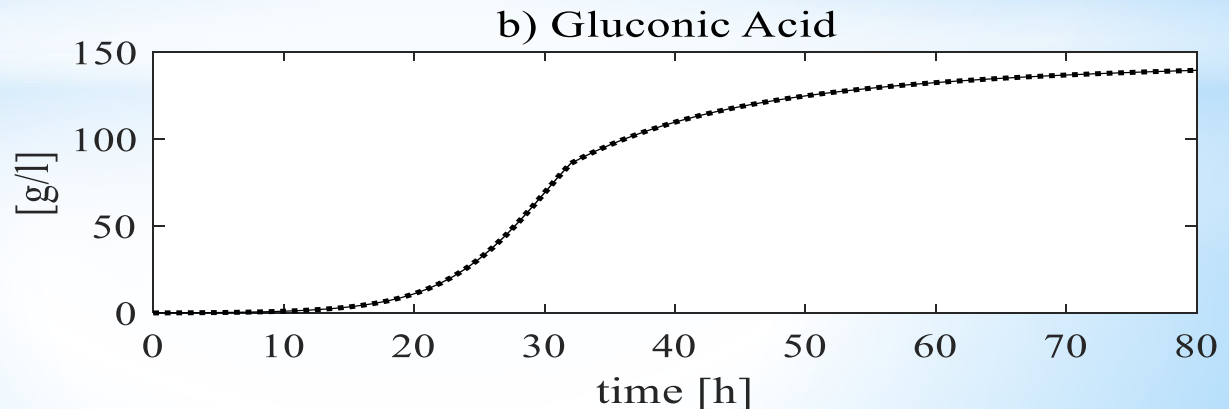
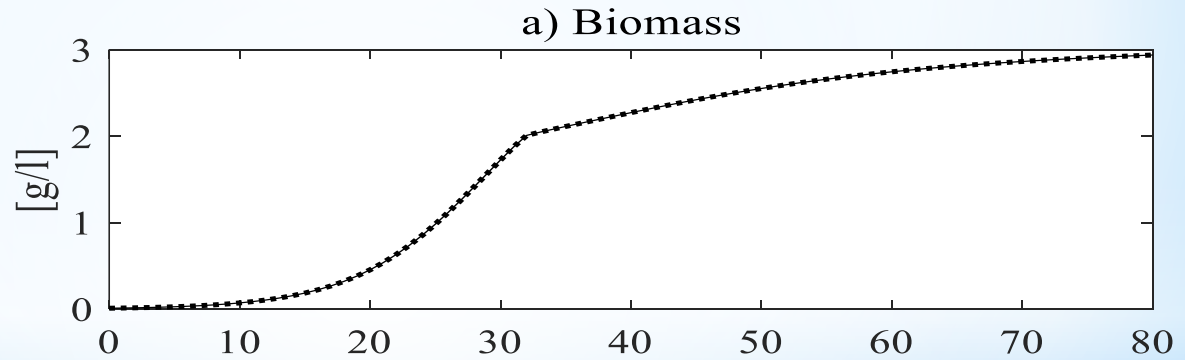


$$X_e = \frac{1}{k_1} Z_1 - \frac{k_2}{k_1 k_3} (Z_2 - O_2) - \frac{1}{k_1} G$$

$$GA_e = \frac{1}{k_3} (Z_2 - O_2)$$

$$\frac{dZ_1}{dt} = -DZ_1 + DG_{in}$$

$$\frac{dZ_2}{dt} = -DZ_2 + K_L a (O_2^* - O_2)$$



Linear regression form of the control model

$$\frac{dX_e}{dt} = X_e G \theta_1 - D X_e$$

$$\frac{dG}{dt} = -X_e G \theta_2 - G O_2 \theta_3 - D(G - G_{in})$$

$$\frac{dO_2}{dt} = G O_2 \theta_5 - D G A_e = -G O_2 \theta_4 - D O_2 + K_L \alpha (O_2^* - O_2)$$

$$\frac{dG A_e}{dt} = G O_2 \theta_5 - D G A_e$$

Continuous Control of **Glucose concentration**

First step $\frac{dG}{dt} = -X_e G \theta_2 - G O_2 \theta_3 - D(G - G_{in})$

Second step $\lambda(G^* - G) = dG/dt$

Third step $D = \frac{-\lambda(G^* - G) - X_e G \theta_2 - G O_2 \theta_3}{G - G_{in}}$

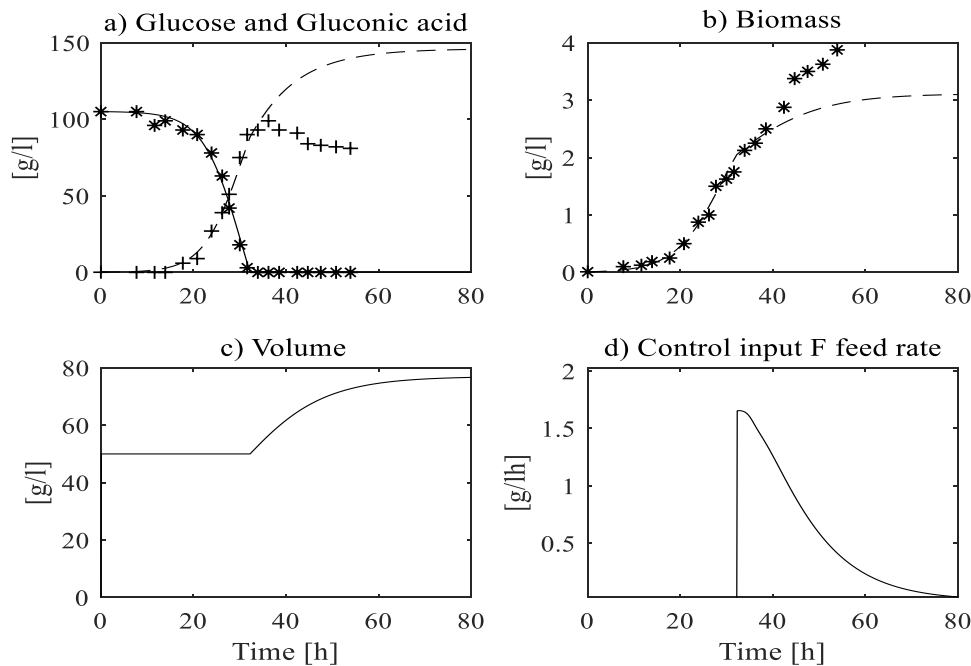
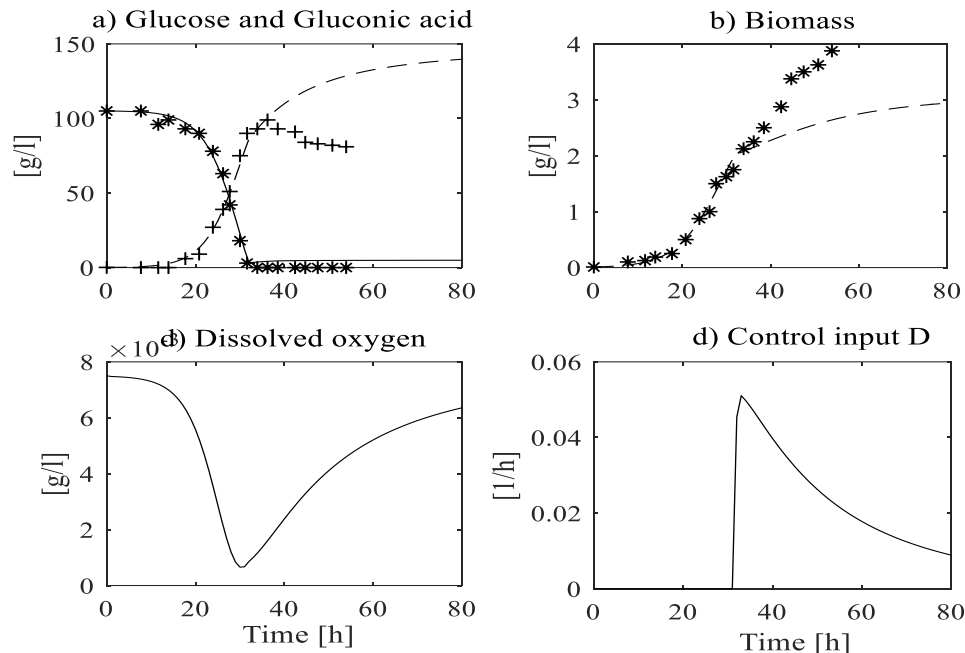
Fed-Batch Control of **Glucose concentration**

First step $\frac{dG}{dt} = -X_e G \theta_2 - G O_2 \theta_3 - \frac{F}{V}(G - G_{in})$

Second step $\lambda(G^* - G) = dG/dt$

Third step $F = \frac{G_{in}(-\lambda(G^* - G) - X_e G \theta_2 - G O_2 \theta_3)}{G - G_{in}}$

$$dV/dt = F$$



Two approaches are considered. The first is to maintain some low value of the substrate (G) in the culture medium (0 or 3 g/l). Comparing the results obtained until the eightieth hour of fermentation in sub-figures a, it was found that a fed-batch control achieves a higher concentration of the target product compared to the continuous control.

Continuous Control of **Gluconic Acid** Concentration

First step
$$\frac{dGA_e}{dt} = GO_2\theta_5 - DGA_e$$

Second step
$$\lambda(GA^* - GA_e) = dGA_e/dt$$

Third step
$$D = \frac{-\lambda(GA^* - GA_e) + GO_2\theta_5}{GA_e}$$

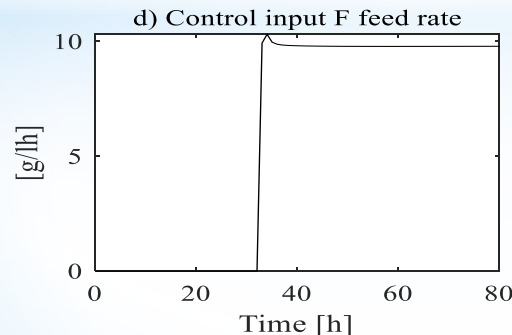
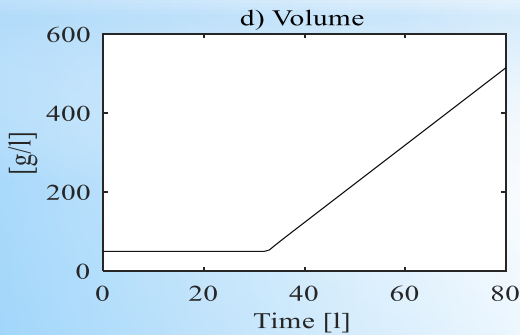
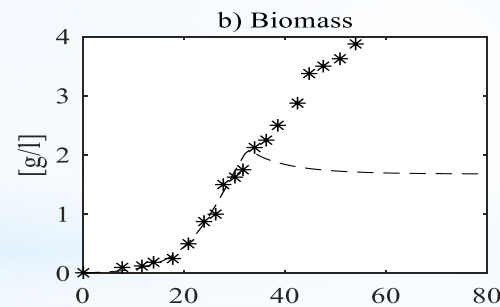
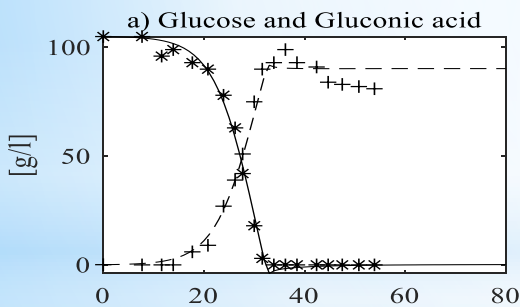
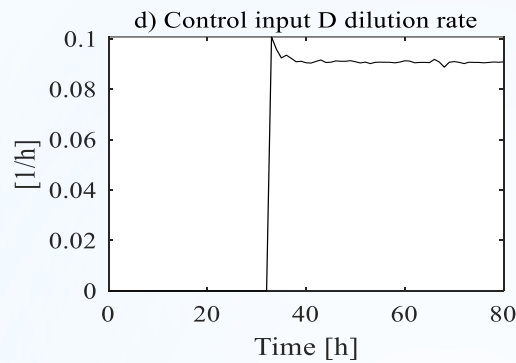
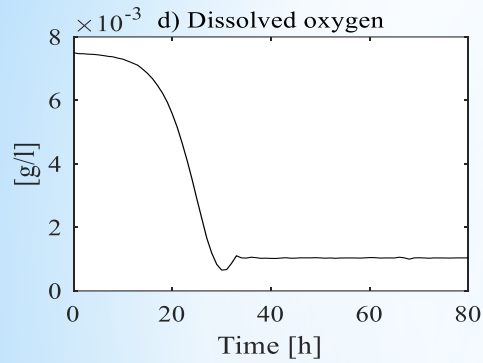
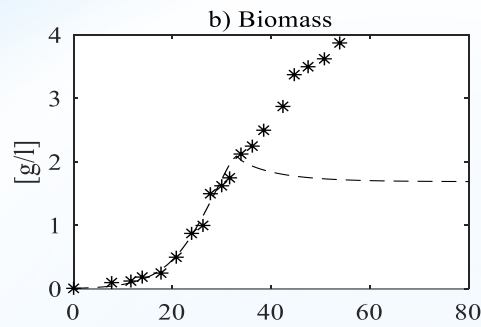
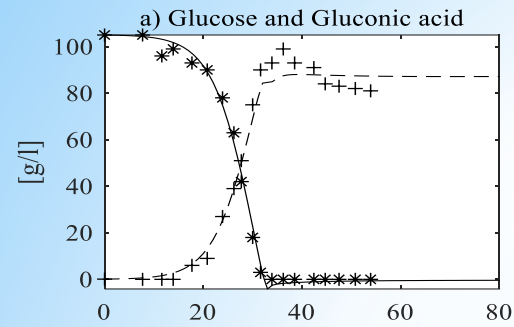
Fed-Batch Control of **Gluconic Acid** Concentration

First step
$$\frac{dGA_e}{dt} = GO_2\theta_5 - \frac{F}{G_{in}} GA_e$$

Second step
$$\lambda(GA^* - GA_e) = dGA_e/dt$$

Third step
$$F = \frac{V(\lambda(GA^* - GA_e) - GO_2\theta_5)}{GA_e}$$

$$dV/dt = F$$



In the second one, a constant concentration of the target product (GA), which corresponds to its maximum productivity is maintained. It should be noted that the fed-batch process must be stopped when the volume reaches the maximum working volume (80 l), while during continuous mode of cultivation, the process may take longer. This does not give grounds for a definite conclusion under which cultivation regime will accumulate a larger amount of target product.

Conclusions

The algorithms developed in this study will be integrated into the system together with similar algorithms, developed for other food production processes. The choice of a process as well as the activation of the functions related to the identification, monitoring and control will be realized by the users through simple actions. In this way, users will be able to get acquainted with the results of modern algorithms without being familiar with the theory of their development, as well as with the software of their implementation. What has been said so far gives grounds to define the system under developing InSEMCoBio as interactive and user-friendly.

**THANKS FOR YOUR
ATTENTION**